

Human Recombinant κ -Opioid Receptor OPRK1 Stable Cell Line

Technical Manual No. TM0442

Version 06042010

I	Introduction.....	1
II	Background.....	1
III	Representative Data.....	2
IV	Thawing and Subculturing.....	2
V	References	3
	Limited Use License Agreement.....	4

I. Introduction

Catalog Number: M00290

Cell Line Name: CHO-K1/OPRK1/G α 15

Gene Synonyms: OPRK1, OPRK, KOR-1, KOR

Expressed Gene: Genbank Accession Number NM_000912; no expressed tags

Host Cell: CHO-K1/G α 15

Quantity: 2 vials (3×10^6 per vial) frozen cells

Stability: 16 passages

Application: Functional assay for OPRK1 receptor

Freeze Medium: 45% culture medium, 45% FBS, 10% DMSO

Complete Culture Medium: Ham's F12, 10% FBS

Culture Medium: Ham's F12, 10% FBS, 100 μ g/ml Hygromycin B, 200 μ g/ml Zeocin

Mycoplasma Status: Negative

Storage: Liquid nitrogen immediately upon delivery.

II. Background

Opioid receptors and their endogenous peptide ligands play important roles in the reward and reinforcement of drugs such as heroin, cocaine, and alcohol. The κ -opioid receptor is a type of opioid receptor which binds the peptide opioid dynorphin as the primary endogenous ligand. κ -opioid receptors are widely distributed in the brain, spinal cord, and in pain neurons. They are associated with the risk for alcohol dependence.

III. Representative Data

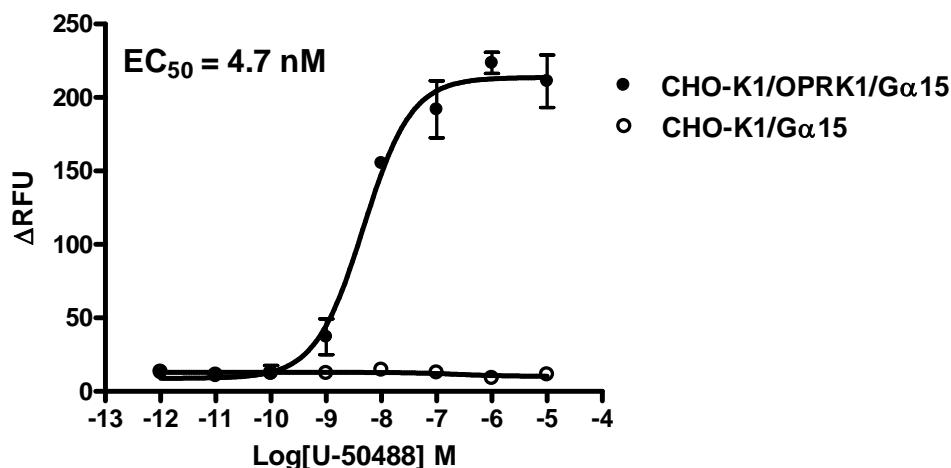


Figure Intracellular calcium response from CHO-K1/Gα15 cells stably expressing human κ opioid receptor OPRK1 and from untransfected control cells. Cells were loaded with Calcium-4 then stimulated with the indicated concentrations of U-50488. Calcium responses were recorded on a FlexStation plate reader. Data represent the average +/- standard deviation of triplicate determinations.

IV. Thawing and Subculturing

Thawing: Protocol

1. Remove the vial from liquid nitrogen tank and thaw cells quickly in a 37°C water-bath.
2. Just before the cells are completely thawed, decontaminate the outside of the vial with 70% ethanol and transfer the cells to a 15 ml centrifuge tube containing 9 ml of complete growth medium.
3. Pellet cells by centrifugation at 200 x g force for 5 min, and discard the medium.
4. Resuspend the cells in complete growth medium.
5. Add 2 ml of the cell suspension per well in a 6 well-plate.
6. Add Hygromycin B and Zeocin to concentrations of 100 µg/ml and 200 µg/ml respectively the following day.

Subculturing: Protocol

1. Remove and discard culture medium.
2. Wash cells with PBS (pH=7.4) to remove all traces of serum that contains trypsin inhibitor.
3. Add 2.0 ml of 0.05% (w/v) Trypsin- EDTA (GIBCO, Cat No. 25300) solution to 10 cm dish and observe the cells under an inverted microscope until cell layer is dispersed (usually within 3 to 5 minutes).
Note: To avoid clumping, do not agitate the cells by hitting or shaking the dish while waiting for the cells to detach. Cells that are difficult to detach may be placed at 37°C to facilitate dispersal.
4. Add 6.0 to 8.0 ml of complete growth medium and aspirate cells by gently pipetting, centrifuge the cells 200 x g force for 5min, and discard the medium.
5. Resuspend the cells in complete growth medium with Hygromycin B and Zeocin and add appropriate aliquots of the cell suspension to new culture vessels.
6. Incubate cultures at 37°C.

Subcultivation Ratio: 1:3 to 1:8 weekly.
Medium Renewal: Every 2 to 3 days

V. References

1. Zhu, J., Luo, L. Y., Li, J. G., Chen, C. and Liu-Chen, L. Y. (1997) Activation of the cloned human kappa opioid receptor by agonists enhances [³⁵S]GTPgammaS binding to membranes: determination of potencies and efficacies of ligands, *J. Pharmacol. Exp. Ther.*, 282, 676 - 684.
2. Chavkin, C., Sud, S., Jin, W., Stewart, J., Zjawiony, J. K., Siebert, D. J., Toth, B. A., Hufeisen, S. J. and Roth, B. L. (2004) Salvinorin A, an active component of the hallucinogenic sage salvia divinorum is a highly efficacious kappa-opioid receptor agonist: structural and functional considerations. *J. Pharmacol. Exp. Ther.*, 308, 1197 - 1203.

GenScript USA Inc.
120 Centennial Ave., Piscataway, NJ 08854
Tel: 732-885-9188, 732-885-9688
Fax: 732-210-0262, 732-885-5878
Email: info@genscript.com
Web: <http://www.genscript.com>

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