

Extraction of Non-Polar Basic Drugs from Plasma with Polymeric SPE Cation Exchange, Bond Elut Plexa PCX

Application Note

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Introduction

Bioanalytical methods for pharmaceutical analysis require quick and easy method development and validation to reduce bottlenecks in drug development. Biological samples can be complicated to analyze due to proteins, peptides, salts, phospholipids and other endogenous compounds. Sample clean-up is necessary to remove these inferences without significant loss of the target analytes. Solid phase extraction utilizing simplified methodologies for routine analysis are the techniques of choice.

Bond Elut Plexa PCX is a new addition to the Plexa family and uses a polymer cation exchange technique. Plexa PCX utilizes a generic and simplified method to remove neutral and acidic interferences from the matrix and concentrate basic analytes resulting in improved analytical performance and sensitivity in the quantitation of basic compounds. In addition, faster and highly reproducible flow rates are the norm, resulting in excellent tube-to-tube and well-to-well performance. Plexa PCX significantly reduces ion suppression because its highly polar, hydroxylated surface is entirely amide-free. The particle exterior excludes proteins and avoids strong binding of phospholipids. Thus, efficient removal of phospholipids from plasma is ensured. A simple generic method was developed for the extraction and analysis of non-polar basic compounds in human plasma.



Materials and Methods

Table 1. SPE Reagents and Solutions

$5\% \text{ NH}_3$ Methanol:acetonitrile (1:1, v/v)	Add 50 μL of concentrated ammonia to 1 mL of methanol:acetonitrile
Methanol:acetonitrile (1:1, v/v)	Add 1 mL of methanol to 1 mL of acetonitrile
2% Formic Acid	Add 20 μL of concentrated formic acid to 1 mL of DI water
Methanol	Reagent grade or better
2% Phosphoric Acid	Add 20 µL of concentrated H ₃ PO ₄ to 1 mL of DI water

Bond Elut Plexa 10 mg 96 well plate (part number A4968010)

Table 2. SPE Method

Sample Pre-treatment	100 µL human plasma. Dilute 1:3 with 2% H ₃ PO ₄ .
Condition	1. 500 μL CH ₃ OH 2. 500 μL DI H ₂ O
Load	Sample with the drug mixture at the flow rate of 1 mL/min
Wash 1	500 µL 2% formic acid
Wash 2	500 μL acetonitrile:methanol (1:1, v/v)
Elution	500 μL 5% NH ₃ methanol:acetonitrile

All samples are evaporated to dryness and reconstituted in 100 μL of 80:20 0.1% Aq formic acid: CH_0H.

Results and Discussion

LC Conditions				
Mobile Phase:	A: 0.1% Fo	rmic acid	l	
	B: Methan	ol		
Gradient:	t = 0 min		80% A : 20% B	
	t = 0-2 min		20% A : 80% B	
	t = 3.5-5 m	in	80% A : 20% B	
Column:	Pursuit C1	8 3 µm, 5	0 x 2.0 mm	
	(part numb	er A3051	050X020)	
MS Conditions	6			
Transition ions	and collision	on energy	/ were:	
Compound	01	03	CE	
Ranitidine	315.0	176.0	-21.0V	
Propranolol	260.1	116.0	-17.5V	
Amitriptyline	278.1	233.0	-17.0V	
Loratadine	383.1	337.0	-31.0V	
Capillary = 25	V, Dry gas t	emp = 4(10 °C, 30 psi,	
CID = Argon				
Polarity:	Positive			



Figure 1. Chromatograms of a 50 ng/mL extract

Table 3. Recoveries of non-polar basic compounds from human plasma

Analyte	log P	рКа	% Rec	% RSD ²	% Rec	% RSD ²	
			(500 ng/mL)		(1000 ng/mL)		
Ranitidine	1.9	8.2	101	5	94	6	
Propranolol	3.6	9.5	97	7	92	4	
Amitriptyline	4.6	9.4	95	5	91	5	
Loratadine	5.2	9.3	100	4	91	4	

 1 Recoveries calculated as % of signal intensity of an extracted sample compared to that calibration curve. 2 RSD = standard deviation/average recovery x 100; n = 6.

This LC/MS method describes the
quantitative determination of non-polar
basic compounds in human plasma
using Bond Elut Plexa PCX for SPE
(Figure 1). The Limit of Detection
(LOD) of the solid phase extraction
and LC/MS/MS analysis was 1.0
ng/mL. Recoveries were calculated
from a 2nd order regression with RSD
values based on a sampling of $n = 6$.
Excellent recoveries were achieved,
demonstrating good retention and
elution, as well as minimal ion
suppression. Response for all the
compounds evaluated was linear
up to 3 orders of magnitude from
1.0 ng/mL to 1.0 μ g/mL with
correlation coefficients all above 0.999.

To demonstrate reproducibility, samples were analyzed at two different concentrations (n = 6). As shown in Table 3, reproducibly high recoveries were obtained according to the generic standard protocol.

Conclusions

With Bond Elut Plexa PCX, it is possible to use a single method for the extraction of non-polar basic analytes from plasma that delivers reproducibly high recoveries. Under acidic conditions, the charged analyte binds to the cation-exchange groups of the sorbent (see Table 3 for pKa). Polar interferences and proteins are washed away with an acidic, aqueous solution. A neutral wash with relatively strong solvents, such as 50% methanol:acetonitrile, is possible without loss of analyte. The wash elutes neutral compounds retained in the hydrophobic cores of the sorbent. Finally, a mixture of organic solvents with ammonia is used to disrupt the cation exchange interaction, resulting in the elution of the basic drugs.

Flow rate over the 96-well plate is fast because Plexa PCX particles have much smaller interstitial paths with no fines to cause blockages, resulting in high well-to-well reproducibility. Automated 96-well technology is convenient which opens new opportunities to maximize efficiency. Bond Elut Plexa PCX is therefore a useful tool for highthroughput SPE applications which require analysis at low analyte levels, need validated reproducibility, and that must be quickly implemented with minimal method development. It is highly recommended for bioanalytical work in pharmaceutical clinical trials, including contract research.

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