

Quantitative Analysis of Water-Soluble B-Vitamins in Cereal Using Rapid Resolution LC/MS/MS

Authors

Sheher Mohsin Agilent Technologies, Inc. Schaumburg, II USA

Michael Zumwalt Agilent Technologies, Inc. 9780 S. Meridian Blvd. Englewood, CO 80112-5910 USA

Indarpal Singh ConAgra Foods, Inc. Omaha, NE 68102 USA

Abstract

An Agilent 6410 Triple Quadrupole Mass Spectrometer (QQQ) is used to analyze several water-soluble B-vitamin compounds in breakfast cereal. A simple gradient elution is carried out on a Rapid Resolution High Throughput SB-Aq column (particle size 1.8 µm). All compounds elute in less than 7.5 minutes, and with the exception of pyridoxine, good linearity over more than three orders of magnitude, from 0.5 to 500 ppb, is demonstrated with good peak area reproducibility at the 0.5 ppb level, which is the lowest level of quantitation considered.

Introduction

Water-soluble vitamins are very polar and have poor retention on reverse-phase columns. The pres-

ence of ion pair reagents such as heptafluorobutyric acid in the mobile phase has been shown to improve the separation and retention of these compounds. However, the drawback of using such ion pair reagents is the high background levels that are generated inside the mass spectrometer. Therefore, we have developed a rapid and sensitive method with ammonium formate in the mobile phase solvent using a column with a bonded phase designed to retain hydrophilic compounds.

The Agilent 1200 Series liquid chromatography (LC) system used in this work was designed to take advantage of sub-2-micron particle columns for rapid, high-resolution separations. Included in the LC design were decreased delay volume, increased pressure range, and increased column temperature. This LC system was coupled to the Agilent 6410 Triple Quadrupole Mass Spectrometer (QQQ) by way of the G1948B electrospray ionization source. Target compound separation was achieved on a ZORBAX AQ 1.8-micron column using a water and methanol gradient with ammonium formate.

Typical LC/MS methods for water-soluble vitamins have shown analysis times as high as 30 minutes with heptafluorabutyric acid ion pairing reagent in the mobile solvent. We have developed a rapid and sensitive method for the LC/MS/MS analysis of water-soluble vitamins by employing a high-efficiency 1.8-micron column in a low-dispersion, 600-bar LC/MS configuration that allowed screening and quantitation with a run-to-run cycle time as low as 10 minutes. Linearity of the mass spec-



trometer response was observed over three orders of magnitude with limits of quantitation in the $0.5 \text{ pg/}\mu\text{L}$ range for all of the analytes except for pyridoxine. In the case of pyridoxine, good sensitivity was demonstrated, but good linearity was limited to just under three orders of magnitude. Calibration curves and chromatograms for the vitamins between 0.5 and 500 pg/ μL were generated for all compounds with the exception of pyridoxine, which was between 0.5 and 250 pg/ $\mu L.$

The structures of the B vitamins are shown below.

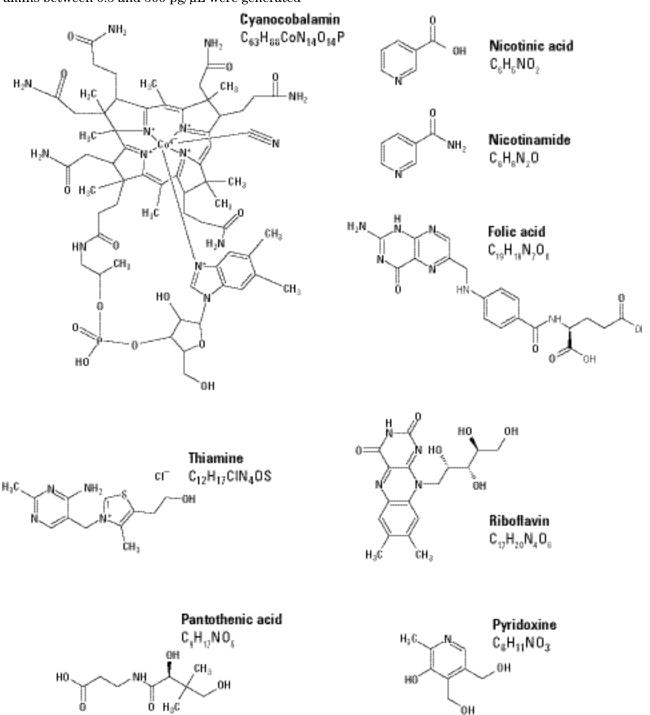


Figure 1. Structures of B vitamins analyzed in this work.

Experimental

Sample Preparation

A standard mix containing all eight compounds in methanol was provided by ConAgra Foods and diluted in 90% water/10% methanol with 20 mM ammonium formate and 0.1% formic acid to the following concentrations: 500, 250, 100, 50, 5, and 0.5 pg/ μ L. These dilutions were used for the quantitation of unknown samples.

One B-vitamin-fortified sample was also provided using the following sample preparation procedure:

- 1. Grind and homogenize breakfast cereal in blender
- 2. Weigh 1 gram of homogenized sample into a 50-mL vial
- 3. Add 25 mL of 0.1M HCl and heat in water bath at 100 °C for 20 min. This solubilizes the vitamins.

- 4. Cool to ambient temperature
- 5. Adjust volume to 1 L with deionized water
- Filter with 0.45-μm glass microfiber membrane.

It should be noted that the provided fortified sample was created for testing the sensitivity of the instrument for customer demonstration purposes. A typical unfortified sample extraction consists of 1 g homogenized sample treated with enzymatic digestion, to release naturally occurring vitamins from their conjugated forms, and volume adjusted to 10 mL, which is 1/100th the volume used in the fortified sample analyzed in this work. At the higher concentration, salts and other matrix contributions are seen to cause interference in the analysis of some vitamins. As a result, further dilution may be used to accommodate the matrix effect in these samples.

Table 1. MRM Mode Parameters

Segment	Compound	Transition	Fragmentor (V)	Collision Energy (V)
1	Thiamine	265.2 > 122.0	85	10
	Pantothenic acid	220.2 > 90.0	110	13
	Pyridoxine	170.1 > 152.1	100	10
	Nicotinic acid	124.1 > 80.0	100	27
	Nicotinamide	123.1 > 80.0	100	25
2	Cyanocobalamin	678.6 > 146.7	130	35
	Folic acid	442.2 > 295.1	120	10
	Riboflavin	377.2 > 243.1	110	25

LC/MS Method Details

LC Conditions

Agilent 1100 Series binary pump, degasser, wellplate sampler, and thermostatted column compartment

Column: Column temperature:	Agilent ZORB 35 °C	AX RRHT	SB-Aq, 3.0 mm × 100 mm, 1.8 μm (PN: 828975-314)				
Mobile phase:		mmonium	formate and 0.1% formic acid in water				
	B = 20 mM al	mmonium	formate and 0.1% formic acid in methanol				
Flow rate:	0.5 mL/min						
Injection volume:	10 µL						
Gradient:	Time (min)	%B					
	0.0	10					
	8.0	55	Stop time: 10 min				
	8.1	10					
Needle wash:	75:25 methanol/water (flush port 20 seconds)						
MS Conditions							
Mode:	Positive ESI u	ising the <i>l</i>	Agilent G1948B ionization source				
Nebulizer:	30 psig	÷					
Drying gas flow:	10 L/min						
Drying gas temperture:	350 °C						
V _{cap} :	1850 V						
Resolution (FWHM):	Q1 = low res; Q2 = low res						
Dwell time for all MRM	200 msec						
transitions							

The precursor ion mass for cyanocobalomin $(m/z \ 678.6)$ is about half of the expected value in which the empirical formula for this compound is $C_{63}H_{88}CoN_{14}O_{14}P$, as denoted in Figure 1. From a correspondence with an analytical chemist (see Acknowledgments) who has run this compound on an Agilent ion trap mass spectrometer, it is shown that the m/z 678.6 actually represents a doubly charged form of cyanocobalomin.

Figure 2A shows the isotopic profile of the ionized cyanocobalomin and since the [13]C isotope contribution is only 0.5 amu higher in mass from the [12]C isotope at 678.1, the profile represents a charge state of 2. Furthermore, in Figure 2B, the full-scan MS/MS of cyanocobalomin is shown with higher mass product ions like m/z 997.5 present. The higher mass product ions are singly charged.

Results and Discussion

The calibration curves for all eight compounds are shown in Figures 3A through 3H. Only for the compound pyridoxine is the 500 ppb level needed to be removed for good linearity. No internal standard is included.

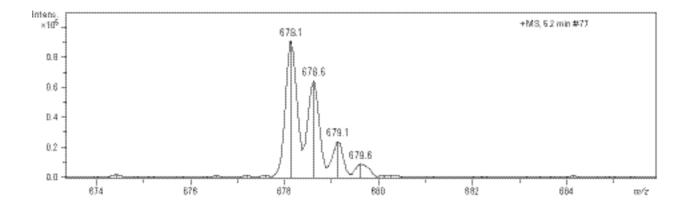


Figure 2A. Doubly charged isotopic profile of cyanocobalomin acquired on Agilent ion trap.

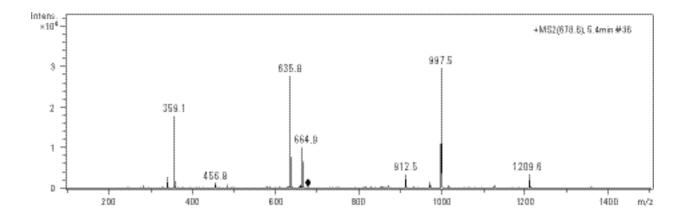


Figure 2B. MS/MS spectrum of the doubly-charged ion of cyanocobalamin.

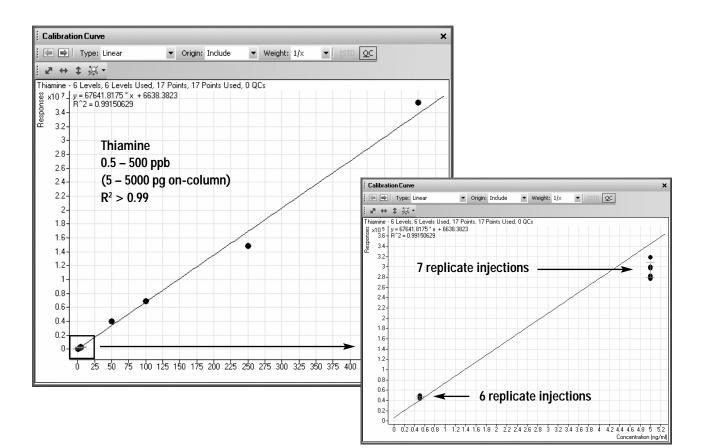


Figure 3A. Linearity of thiamine over three orders of magnitude.

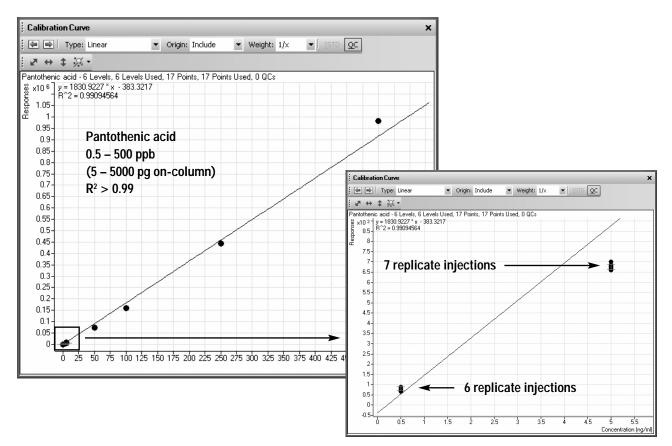


Figure 3B. Linearity of pantothenic acid over three orders of magnitude.

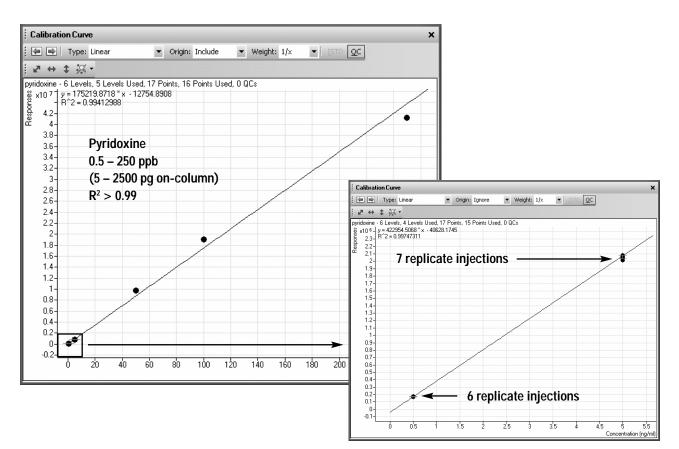


Figure 3C. Linearity of pyridoxine acid over nearly three orders of magnitude.

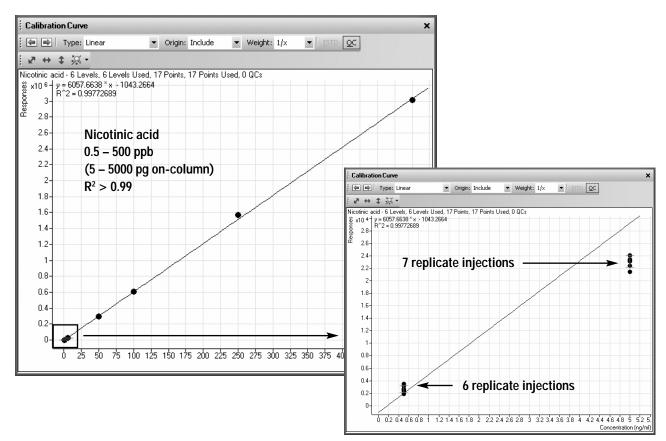


Figure 3D. Linearity of nicotinic acid over three orders of magnitude.

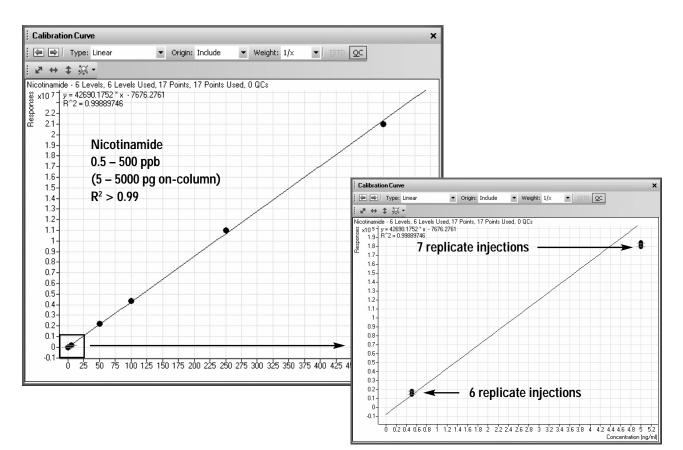


Figure 3E. Linearity of nicotinamide acid over three orders of magnitude.

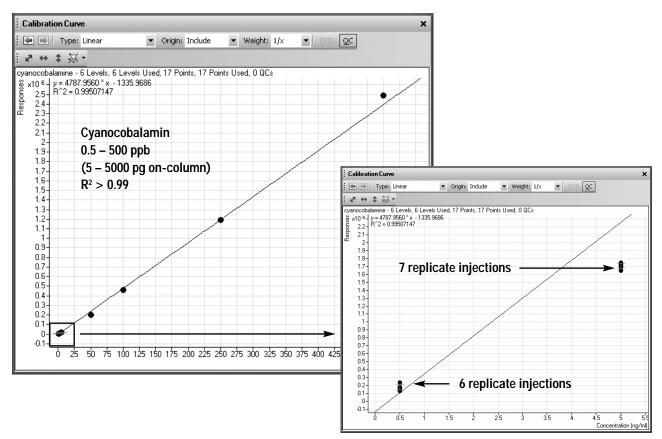


Figure 3F. Linearity of cyanocobalamin over three orders of magnitude.

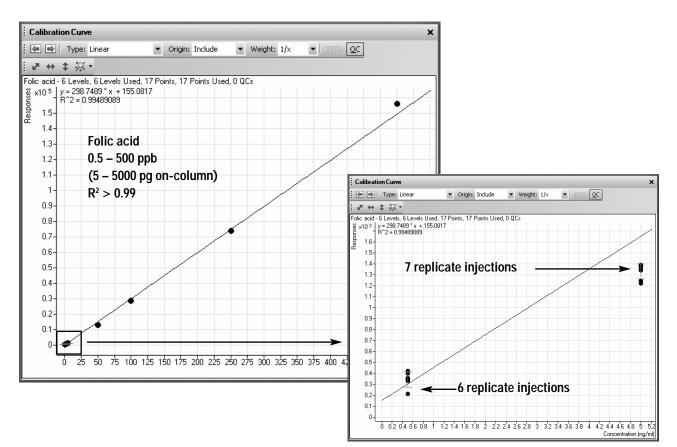


Figure 3G. Linearity of folic acid over nearly three orders of magnitude.

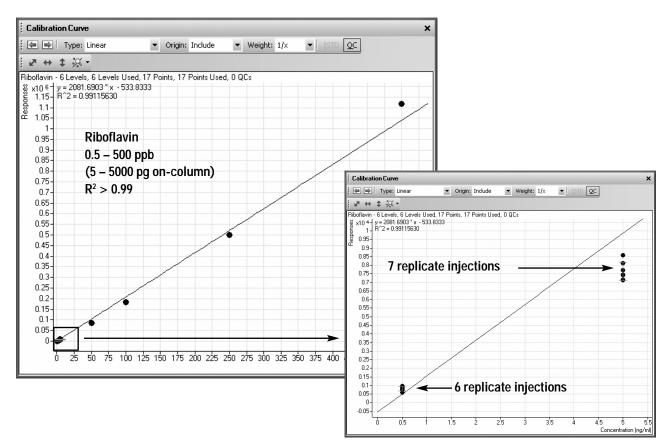


Figure 3H. Linearity of riboflavin acid over three orders of magnitude.

All line fits to data are carried out as linear with the origin ignored and a 1/x weighting.

An example of reproducibility at the 0.5 ppb level for pyridoxine is shown in Figure 4. The peak area %RSD values for all compounds at the 0.5 and 5 ppb level are given in Table 2.

Table 2.	Peak Area Reproducibility for Each Compound at the
	Two Lowest Levels Used for Quantitation

	Level %RSD						
Compound	0.5 ppb	5 ppb					
Thiamine	5.9	1.8					
Pantothenic acid	14.0	3.3					
Pyridoxine	1.7	1.0					
Nicotinic acid	9.2	1.8					
Nicotinamide	5.5	0.7					
Cyanocobalamine	2.7	1.3					
Folic acid	20.0	3.9					
Riboflavin	4.4	2.7					

A fortified cereal extract is also analyzed and quantitated using the diluted standard mix already mentioned. An example of the batch results using the MassHunter Quantitative Analysis is shown in Figure 5. The concentration of nicotinamide present in the sample is calculated to be $116.2 \text{ pg/}\mu\text{L}$.

The concentrations calculated for all compounds in the fortified extract are given in Table 3.

The corresponding chromatographic elution of the eight compounds detected in the fortified extract is shown in Figure 6.

 Table 3.
 Calculated Concentrations for Each Compound in Fortified Cereal Extract

	Calculated
	concentration
Compound	(pg/µL)
Thiamine	24.0
Pantothenic acid	1.2
Pyridoxine	15.5
Nicotinic acid	43.5
Nicotinamide	116.2
Cyanocobalamin	0.4*
Folic acid	2.6
Riboflavin	8.6

* Outside quantitation limits.

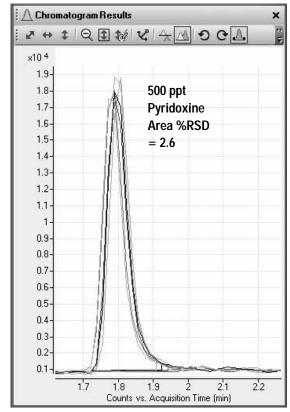


Figure 4. Peak area reproducibility of pyridoxine at 500 ppt level, 6 injections.

San	ple:	1 2	Sample Ty	/pe: <a< th=""><th>👻 Compound: 💽</th><th>1: Nicotinar</th><th>nide</th><th>• 🖻 1</th><th>STD:</th><th>TI</th><th>ne Segi</th><th>nen</th><th>C <a -="" th="" 🗔<=""><th>1 R</th><th>24</th></th></a<>	👻 Compound: 💽	1: Nicotinar	nide	• 🖻 1	STD:	TI	ne Segi	nen	C <a -="" th="" 🗔<=""><th>1 R</th><th>24</th>	1 R	24
				Sam	pic			Nicotina	Nicotinamide Results						
۲	8	Name	Туре	Level	Acq Date-Time /	Data File	Dii	Exp. Conc.	RT	Resp.	S/N	MI	Calc Conc	Final Conc.	Accurac
T	14	bink	Blank	1	1/26/2007 1:23 PM	blnk1.d	1.0		3.306	507	0.12		0.0299	0.0299	
		0.5ppb	Cal	1	1/26/2007 2:55 FM	500ppt1.d	1.0	0.5000	3.245	45440	14.51		0.4955	0.4955	99.
		0.5ppb	Cal	1	1/26/2007 3:07 PM	500ppt2.d	1.0	0.5000	3.251	41164	12.48		0.4512	0.4512	90.
		0.5ppb	Cal	1	1/26/2007 3:18 PM	500ppt3.d	1.0	0.5000	3.236	45058	14.15		0.4916	0.4916	98
		0.5ppb	Cal	1	1/26/2007 3:30 PM	500ppt4.d	1.0	0.5000	3.260	44073	14.74		0.4814	0.4814	96.
		0.5pph	Cal	1	1/26/2007 3-41 PM	500ppt5 d	10	0.5000	3 245	42910	16.45		0.4693	0.4693	93
1		0.5ppb	Cal	1	1/26/2007 3:53 PM	500ppt6.d	1.0	0.5000	3.249	39420	12.32	1	0.4332	0.4332	86.
		5ppb	Cal	2	1/26/2007 4:04 PM	5ppb1.d	1.0	5.0000	3.255	488810	61.80	4	5.0900	5.0900	101
		5ppb	Cal	2	1/26/2007 4:16 PM	5ppb2.d	1.0	5.0000	3.238	483252	70.05	2	5.0324	5.0324	100.
		5ppb	Cal	2	1/26/2007 4.27 PM	5ppb3.d	1.0	5.0000	3.258	494014	76.35		5.1439	5 1439	102.
		5ppb	Cal	2	1/26/2007 4:39 PM	5ppb4.d	1.0	5.0000	3.248	488774	78.38		5.0896	5.0896	101.
		5ppb	Cal	2	1/26/2007 4:50 PM	5ppb5.d	1.0	5.0000	3.257	489640	83.64		5.0986	5.0986	102
		5ppb	Cal	2	1/25/2007 5:02 PM	5ppb6.d	1.0	5.0000	3.259	486089	79.07		5.0618	5.0618	101.
		5ppb	Cal	2	1/26/2007 5:13 PM	5ppb7.d	1.0	5.0000	3.250	484797	76.87		5.0484	5.0484	101.
		50ppb	Cal	3	1/25/2007 5:25 PM	50ppb.d	1.0	50.0000	3.255	5413252	52.58	\bigtriangledown	56.1193	56.1193	112.
	1	100ppb	Cal	4	1/26/2007 5:36 PM	100ppb.d	1.0	100.0000	3.255	10915097	77.91		113.1320	113.1320	113.
		250ppb	Cal	5	1/25/2007 5:48 PM	250ppb.d	1.0	250.0000	3.255	25481085	84.13	\square	264.0/16	264.0/16	105.
0		500ppb	Cal	6	1/26/2007 5:59 PM	500ppb.d	1.0	500.0000	3.255	44995630	573.69	4	466.2904	466.2904	93.
		blnk	Blank	1	1/26/2007 6:22 PM	blnk3.d	1.0	()		9	1 - E		3		Į
		sample	Sample		1/26/2007 6:57 PM	sample1b.d	1.0		3.258	11213094	93.72		116.2200	116.2200	

 $\label{eq:Figure 5. Concentration calculated to be 116.2 \ pg/\mu L \ (highlighted).$

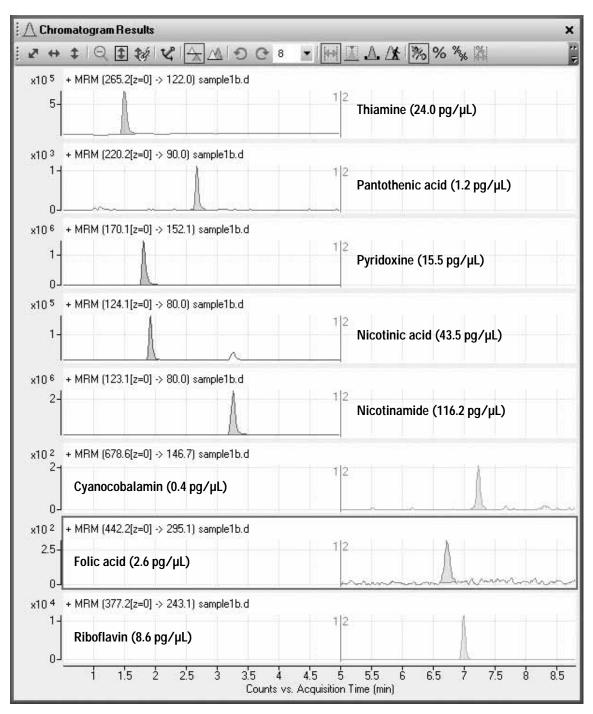


Figure 6. Chromatogram of compounds in fortified extract and calculated concentrations.

Conclusions

The water-soluble B vitamins are successfully analyzed using LC/MS/MS. Good linearity with at least $R^2 > 0.99$ is demonstrated over three orders of magnitude for all compounds, with reproducibility as low as 1.7 %RSD at the lowest level of quantitation for pyridoxine. An extracted fortified sample is successfully analyzed with only the cyanocobalamin concentration falling below the quantitation limit.

Acknowledgements

The authors gratefully acknowledge the assistance of Melissa Medrano Gomes, Universidade de São Paulo, Faculdade de Ciências Farmacêuticas, Departamento de Análises Clínicas e Toxicológicas, Avenida Professor Lineu Prestes 580 Bloco 17, Cidade Universitária 05508-900, Sao Paulo, Brasil, in identifying the charge state of cyanocobalamin.

For More Information

For more information on our products and services, visit our Web site at www.agilent.com/chem.

For more details concerning this application, please contact Michael Zumwalt at Agilent Technologies, Inc.

> Agilent shall not be liable for errors contained herein or for incidental or consequential damages in connection with the furnishing, performance, or use of this material.

> Information, descriptions, and specifications in this publication are subject to change without notice.

© Agilent Technologies, Inc. 2008

Printed in the USA August 28, 2008 5989-7084EN

