

Agilent PL-SAX HPLC Columns

Data Sheet

The PL-SAX (strong anion) exchange media has an optimized pore size and structure for the analysis of biological macromolecules.

For optimum performance and lifetime please read this data sheet before commissioning and using the column.

Installation

Microbore, analytical, and semi-prep columns up to 25 mm id

A 1/16 in stainless steel tubing is recommended for column connections, 0.002 in id for microbore, 0.010 in id for analytical work and 0.020 in id for preparative work. Connecting tubing lengths should be minimized to avoid excessive dead volume which will diminish system performance. Column connections should be made using Parker compatible 1/16 in nuts and ferrules with special reference to compatibility of column connectors as illustrated in Figure 1. Connect the HPLC column in the flow direction indicated. The nut and ferrule should be tightened 1/4 of a turn past finger tight by applying the wrenches as shown in Figure 2.



Figure 1. Compatible connectors.

The distance "X" for the standard PLRP-S column end fitting is 0.090 in and a minimum male nut length of 0.210 in is required. Some fittings from other manufacturers may not be compatible, for example, Waters and Rheodyne. If unsure, please contact Agilent Technologies.

To avoid loosening the endfittings and causing leaks, wrenches must be used on the endfitting adjacent to the connecting nut and NOT on the column barrel or the opposite endfitting.

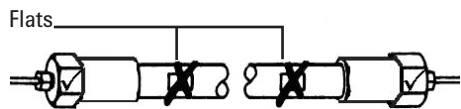


Figure 2. Don't use wrenches on the flats.

Preparative columns 50 and 100 mm id

A 1/16 in or 1/8 in stainless steel tubing is recommended for column connections to the largest preparative columns. The size chosen should be appropriate to the flow rate used and the pressure of the column. Column connections should be made using fittings that are compatible with Vici Valco nuts and ferrules. 1/16 in connections should be made using the reducing unions supplied with the column. Please ensure that the column is secured firmly to the bench before use.

Shipping Eluent

PL-SAX columns are supplied containing 0.1 M Na₂SO₄ and 0.02% sodium azide. Columns are securely sealed with end caps which must always be replaced when the column is disconnected from the system to prevent the columns drying out.

Column Conditioning

It will be necessary to wash out the shipping solution and condition with the required counter ion prior to use. The following procedure is recommended at 180 cm/h (0.5 mL/min for a 4.6 mm id column).

1. Elute for 10 minutes with the low ionic strength component of the mobile phase, buffer A, for example 0.01 M Tris HCl, pH 8.0.
2. Exchange the counter ion by eluting with the high ionic strength component of the mobile phase, buffer B, for example 0.01 M Tris HCl, 0.5 M NaCl, pH 8.0. Continue with this eluent until a stable baseline is achieved at the required sensitivity, a minimum of 10 minutes.
3. Equilibrate with buffer A for a minimum of 2 column volumes prior to use.

Mobile Phases

The PL-SAX matrix, being polymeric and macroporous, is stable in most polar mobile phases. The excellent chemical resistance of both the base polymer and of the anion exchange functionality enables the use of aqueous buffers in the pH range 1–13 without accelerated column degradation or loss of ionic capacity.



Agilent Technologies

Both cationic and non-ionic detergents can be used. However, the column will need conditioning with the required detergent prior to use. As an anion exchange column, it is recommended that anionic detergents are not used.

The pre-packed columns are stable in alcohols (C_1 – C_4), however when changing mobile phases, the miscibility and solubility of all eluent components must be examined.

Flow Rate/Pressure

The maximum operating pressure for the PL-SAX stainless steel HPLC column is 3000 psi (207 bar). With low viscosity mobile phases, this will enable flow rates of up to 4.0 mL/min to be used with short 4.6 mm id columns. If column pressures are high, due to mobile phase viscosity, or to improve sample solubility or resolution elevated temperatures 80 °C, can be used.

Preparation

The samples should be free from fat, which would otherwise contaminate the column, and be filtered (< 0.5 μ m). If turbid sample solutions are injected, even after being filtered, the lifetime of the column may be significantly reduced.

If possible the samples should be dissolved in buffer A, the low ionic strength component of the mobile phase. For interaction to occur with the strong anion exchanger the solutes must be negatively charged at the analysis pH. In the case of proteins where the total net charge is pH dependant, this will be above the pI, iso-electric point. The pH can be controlled by the use of any of the commonly used cationic buffers, for example, Tris, piperazine or diethanolamine. The solutes can be eluted by increasing the ionic strength or changing the mobile phase pH.

Column Efficiency Testing

Each column is provided with its own individual test certificate. Agilent recommends that the column be re-checked from time to time to monitor its performance.

Column Clean-Up

If the column starts to exhibit signs of deterioration such as a loss of resolution, increased back pressure or loss of solute recovery, then regeneration is required. When the performance has deteriorated then the contamination will have occurred at the column inlet, therefore the flow through the column should be reversed during washing.

The excellent chemical stability of the PL-SAX media enables washing with 1 M acid, for example, acetic or hydrochloric and 1 M base, for example, sodium hydroxide, to be accomplished. If the contamination is due to small hydrophobic molecules, for example, fats, and detergents, then the matrix should be washed with an organic alcohol for example, isopropanol. The addition of 0.1% trifluoroacetic acid to the organic may be advantageous. After each washing sequence it is recommended that a high salt elution be carried out. After thorough cleaning, the column should be conditioned as detailed earlier.

Storage

On removing the column from the system, the end plugs must be replaced to prevent the columns from drying out by evaporation since disruption of the packed bed may occur. The end plugs should be finger tight only.

For long-term storage, the column should be washed with 1 M sodium chloride. After flushing with water the storage buffer of approximately 0.1 M Na_2SO_4 containing 0.2% sodium azide can be introduced.

Agilent Ordering Information

For more information on our products, visit our web site at www.agilent.com/chem/columns.

www.agilent.com/chem

Agilent shall not be liable for errors contained herein or for incidental or consequential damages in connection with the furnishing, performance, or use of this material.

Information, descriptions, and specifications in this publication are subject to change without notice.

© Agilent Technologies, Inc., 2011
Printed in the USA
February, 2011



PK013



Agilent Technologies