INSTRUMENTS AT WORK

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Applications of UV-Visible Derivative Spectrophotometry

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Part II Some potential areas of application of UV-Visible derivative spectroscopic techniques.

Introduction

In part I, after a very brief historical introduction, an overview of the areas of application was presented, and the basic principles of the derivative technique were discussed in some detail.

The accelerating interest in the use of derivative techniques in UV-Visible spectroscopic analyses is further illustrated by the additional number of recently published papers, some of which are listed here.

The major part of this paper will be devoted to illustrating the power and usefulness of the derivative technique in various areas of UV-Visible spectroscopic measurements.

The various topics in Part II are discussed under the following headings:

Very Recent Areas of Application Clinical-Pharmaceutical-Biochemical (Life Sciences) Inorganic Miscellaneous Experimental Measurements and Discussion Characterization of Individual Pure Compounds Study of Homologous and Isomeric Series of Compounds Quantitative Determination of Trace Components Minimization and Elimination of Background Absorption Characterization of Commercial Materials and Natural Substances Conclusion

Very Recent Areas of Application

In this, again by no means complete, selection of papers published in the last few years the increasing recognition of the usefulness of derivative techniques is further re-inforced, particularly in the so-called 'Life Sciences' area, where analyses most often have to be carried out under adverse conditions, i.e. in the presence of strongly interfering (absorbing and scattering) background matrices.

Clinical-Pharmaceutical-Biochemical (Life Sciences)

Analysis of colouring agents in pharmaceuticals by derivative ultraviolet-visible spectroscopy.	[122]
Determination of morphine and heroin by second derivative UV-Spectrophotometry.	[102]
Pharmaceutical applications of computer-aided optical multi-channel Spectroscopy.	[123]
Stability of oral vitamin K — a comparison of an HPLC and derivative spectrophotometric method	[124]
Application of difference and derivative ultra-violet spectrometry	[120]
First derivative spectrophotometric determination of certain drugs in two-component mixtures	[120]
Evaluation of dual-wavelength spectrophotometry for drug	[127]
Application of first-derivative spectrophotometry to	[128]
component dosage forms.	[129]
Determination of aspirin and salicylic acid in aspirin tablets by second-derivative ultra-violet spectrometry	[130]
Ultra-violet derivative spectrophotometric determination of Cui Xing Ning tablets	[100]
Derivative spectrophotometry and its application in pharmaceutical analysis	[131]
Application of derivative spectrometry in pharmaceutical analysis. II. Determination of guaiphenesin and isoprenaline hydrochloride in aerosol by second-derivative spectrometry	[132]
and colorimetry.	[133]
tablets by second-derivative spectrometry.	[134]
of carbonylhaemoglobin in blood.	[135]
Determination of carbonylhaemoglobin in the presence of other blood haemoglobin pigments by visible spectrophotometry.	[136]
Determination of certain drugs in multi-component formulations by first-derivative ultra-violet spectrophotometry.	[137]
Determination of salicylic acid in aspirin by first-derivative ultra-violet spectrophotometry.	[138]
Atropine sulphate analysis by derivative spectroscopy or HPLC.	[139]
spectrophotometry.	[140]
Determination of coloured substances in soya-bean lecithin (phosphatidylcholine).	[141]
First derivative spectrophotometric determination of pyridoxine and meclozine in two-component mixture.	[142]
Derivative spectrophotometric determination of praziguantel in tablets.	[1/2]
•	[140]

Studies on derivative spectrophotometry. I. Theoretical analysis of factors in the resolution of overlapping	
absorption bands by use of derivative spectrophotometry.	[169]
Effect of the degree of polynomials in the Savitzky-Golay	
method for calculation of second-derivative spectra.	[170]
Quantitative analysis by derivative electronic spectroscopy.	[171]
Application of derivative spectrophotometry to the study	
and analysis of complex substances in solution.	[172]
Determination of alkyInaphthalenes in petroleum fractions	
by second-derivative ultra-violet spectrophotometry.	[173]
Arson analysis by second-derivative ultra-violet spectrometry.	[174]
Derivative spectrophotometry (a literature review).	[175]
Ratios of first-derivative maxima and compensated derivative	
absorption curves.	[176]

Experimental

All spectrophotometric measurements presented in this paper were carried out on a new, recently introduced, microprocessor controlled, double-beam UV-Visible scanning spectrophotometer, the Varian DMS 200 [177], equipped with a BMC monitor and a Sekonic S-210 GP thermal printer-plotter. Details of the measurement parameters, chemicals and solvents are given with the corresponding spectral traces. In all cases, unless specifically indicated otherwise, 1-cm pathlength quartz cells were employed.

The DMS 200 spectrophotometer was chosen for this work primarily for its built-in 1st to 6th derivative measurement and display capabilities. It must be noted, however, that many of its other operational and performance characteristics also played a significant, interactive role in the derivative measurements presented here. Some of these interactions and their impact an measured spectral data are discussed in more detail in this section.

Double-beam operation in the DMS 200 is achieved by means of 3-segment 30 Hz rotating choppers, which ratio the sample and reference signals every 33 ms, with a sampling time of only 11 ms between the sequentially ratioed sample and reference signals. The 3-segment design also provides automatic dark current compensation every chopper cycle (i.e. compensation of the statistical background signal produced when no light falls on the photomultiplier detector).

It should be noted that this sequential signal and reference sampling time in single-beam spectrophotometers is usually of the order of tens of seconds, or even longer, and is dependent on the speed of the human operator. Also, most single-beam instruments have no provision, either manual or automatic, for dark current compensation. Furthermore, the indiscriminate, automatic use of microprocessor stored reference baselines (even on the sample cell with solvent) for subsequent sample spectra corrections can lead to unrecognised, incorrect results (quantitatively and qualitatively). This is especially the case for small signal-to-noise ratio (S/N) peaks and shoulders, and particularly in derivative measurements, because single-beam instrument stabilities, even after a 1-hour warm-up period, are generally 6 to 20 times worse than for double-beam instruments.

For example, Figure 23 shows the double-beam DMS 200 stability (at 500 nm, 2 nm spectral band width and 'zero' smoothing time). Figure 24 shows the stability of the DMS 200 operated in the single-beam mode, but with the advantage of automatic dark current compensation.



Figure 23



Figure 24

Nevertheless, the latter shows some drift and highlights the potential danger of storing a single-beam instrument baseline and then using it for correction on sample measurements performed minutes or hours later.

The DMS 200, like many modern UV-Visible spectrophotometers, has a specified wavelength range of 190 to 900 nm. However, on any instrument the usability of the whole wavelength range will depend on many factors, such as:

The overall optical design and its light throughput efficiency.

The performance of the light sources and the detector.

The slit spectral bandwidth (S.B.W.).

The stray light level (the amount of unwanted radiation which reaches the detector at wavelengths other than that being measured).

The type of sample, solvent and the type of cells being used.

The region below 220 nm may present some difficulties on all spectrophotometers, because of increasing levels of stray light, decreasing light throughput and decreasing detector response, which is aggravated by the increasingly strong absorption of atmospheric oxygen, particularly below 200 nm (Figure 25), and the very strong absorption by most organic substances, including many useful solvents.



Figure 25

However, the low stray light of the DMS 200 plus the excellent energy throughput, even at narrow S.B.W. settings result in low noise and enable measurements to be made successfully in this 'difficult' region, as illustrated by the spectrum of cyclopentanone vapor, measured with 0.2 nm S.B.W. slits (Figure 26).





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In such a spectrum the peaks and shoulders can be further resolved and more clearly identified by the use of higher derivatives. For example, the progressively improved resolution of the 3 peaks in the region of 198 nm to 203 nm is clearly shown by the 2nd and the 4th derivative traces in Figures 27 and 28 respectively.









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The asymmetrical peaks at 200.6 nm and 199.6 nm are doublet peaks.

Wavelength accuracy and wavelength repeatability also play an important role, particularly in spectral characterization and for spectral comparisons of pure substances and for subsequent archiving. Generally, small deviations in wavelength accuracy are to be expected and are not a serious handicap, since such deviations can be readily measured and corrected for, as long as they are constant over the entire wavelength range of interest.

The 656.10 nm and 486.00 nm emission lines from the deuterium arc UV light source, found in most instruments, are particularly useful for routine, fast checks of wavelength accuracy [178].

On the DMS 200 the wavelength accuracy routine check can be made quickly, using the built-in deuterium arc source in the single beam energy mode, with the automatic source change programmed to occur above 656.1 nm (660 to 700 nm for example). A typical routine wavelength accuracy calibration check is shown in Figures 29 and 30.



Figure 29



Figure 30

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An additional benefit obtained from such a check is an indication of the approximate resolution achieved with the selected S.B.W. That is, the band width at peak-half-height gives a good indication of the spectral slit width.

Wavelength repeatability is probably an even more critical parameter for ensuring reproducible photometric measurements, particularly in quantitative work at fixed wavelengths, ideally at peak maxima, but often on the sides of peaks.

In instruments, such as the DMS 200 and Cary Spectrophotometers, improved wavelength reproducibility is due to the selected wavelength always being approached from the same direction (from longer wavelengths), thus minimizing mechanical backlash effects.

This is particularly important as the scanning speeds of UV-Visible spectrophotometers become faster and faster. Of course, the foregoing remarks apply equally well to derivative measurements, but here performance criteria such as noise play an even more critical role.

Noise will be very much dependent on the chosen working slit S.B.W., and inevitably the latter plays a decisive role in spectral peak resolution. As a rough rule-of-thumb, the S.B.W. should be about one-tenth or less of the 'true' or natural bandwidth of a peak so that the error in the peak absorbance is kept to less than 0.5 % [179]. The level of noise in the original zeroth order absorbance spectrum, which increases with decreasing S.B.W., will be of paramount importance for subsequent derivative calculations — the limiting factor for higher order derivative usefulness.

The completely variable S.B.W. setting on the DMS 200 (from 0.2 nm to 4.0 nm), provides a fairly wide range of control over both resolution and noise.

Further noise control is provided through a built-in proprietary noise filter, which is operator accessible via the selection of one of three digital smoothing times for derivative calculations (0.2, 1, and 5 seconds).

Finally, discrimination against noise peaks is provided by the peak threshold facility which enables the operator to select only those peaks which are of significance for printout.

The interplay of these operation parameters is illustrated by, for example, the absorbance and second derivative spectra of a broad peak (Figure 31), where the 1-second smoothed 2nd derivative trace minimum shows excellent wavelength agreement with the 'zero' smoothed absorbance trace maximum.

Trace A Operator Sample Mode Abs SBW (nm) 2.0 Smoothins (sec) 1.000 Ord Max/Min 0.000 WL Max/Min (nm) 350.0 250.0 Speed (nm/min) 100 3 Peaks, threshold 0.200 Min 349.6 nm, 0.011 Max 301.0 nm, 0.527 Min 263.1 nm, 0.132 Trace в Operator Sample 2ND Der Mode SBW (nm) 2.0 Smoothing (sec) 0.2 Ord Max/Min 0.010 -0.010 WL Max/Min (nm) 350.0 250.0 Speed (nm/min) 100 14 Peaks, threshold 0.005 Max 329.6 nm, 0.003 Max 312.0 nm, -0.001 Min 310.8 nm, -0.007 Max 309.8 nm, 0.002 Min 302.9 nm, -0.007 Min 300.6 nm, -0.006 Min 295.8 nm, -0.004 Min 289.0 nm, -0.003 Min 274.2 nm, -0.002 261.5 nm, 0.004 Min Min 259.3 nm, 0.005 Min 255.5 nm, 0.016 Min 253.6 nm, 0.023 Max 251.6 nm, 0.049 Trace С Operator Samele Mode 2ND Der SBW (nm) 2.0 Smoothing (sec) 1 Ord Max/Min 0.010 -0.010 WL Max/Min (nm) 350.0 250.0 Speed (nm/min) 100 3 Peaks, threshold 0.005 Max 325.5 nm, 0.003 Max 252.9 nm, 0.021 Min 301.1 nm, -0.004





The factors which affect derivative measurements can be summarized as:

The type of sample, i.e. having sharp or broad spectral features, requiring narrow or wide S.B.W.'s, respectively.

The sample concentration, which will determine the absorbance level, and therefore produce high or low noise situations.

The wavelength region in which the sample absorbs, again producing either high or low noise conditions. The selectable instrument (DMS 200) operating parameters which will determine the quality of derivative data are:

The scanning speed, which will determine the data sampling interval ($\delta\lambda$) for derivative calculations:

Scan Speed (nm/min)	δλInterval (nm)
20	0.2
50	0.5
100	1.0
200	2.0
500	5.0
1000	10.0

The slit S.B.W., which will determine the resolution and the level of noise:

from 0.2 nm to 4.0 nm, selectable in 0.1 nm steps.

The smoothing filter times, which will determine the number of collected data points taken into the calculation of each derivative point:

Selectable digital smoothing filter times (0.2, 1, and 5 seconds).

Thus, as a rough guide, the following 'trading rules' for the optimization of derivative measurements on the DMS 200 can be put forward:

Narrow Peaks:

For optimum resolution a narrow slit S.B.W. between 0.2 nm — 0.5 nm, should be used, together with a relatively slow scanning speed, ideally 20 nm/min or 50 nm/min, and definitely not faster than 100 nm/min. A 1-second smoothing filter is generally the most appropriate.

Broad Peaks:

Relatively wide slit S.B.W.'s, 1 nm — 4 nm, should be used for minimization of noise, together with medium smoothing filter time (1 or 0.2 second), and medium scanning speeds of 100 nm/min or 200 nm/min for peak amplitude amplification. Unless a broad peak is a composite of several overlapping narrow peaks, going above the 2nd derivative may often prove to be highly questionable.

In general, for rapid survey scans, that is, fast scan speeds of 500 nm/min or 1000 nm/min (large $\delta\lambda$ steps), over a wide wavelength range, derivative measurements are a rather futile exercise.

These easily understandable and controllable optimization parameters, the visual display of spectra on a medium-high resolution CRT screen, together with the extensive on-screen spectral manipulation facilities, prior to print-out on a high-resolution graphics printer-plotter, make the DMS 200 eminently suitable for both normal and derivative UV-Visible spectrophotometric measurements.

Measurements and Discussion

In this section are presented various examples of derivative measurements which illustrate a few of the areas of application of the technique.

Characterization of Individual Pure Compounds

Very often the UV-Visible spectrophotometric technique, on its own, has not been very useful for the characterization of substances, even when pure, and particularly in solutions. The relative non-specifity has hindered its wide application to qualitative analyses. However, the advent of and improvements in derivatization methods have brought new possibilities for the universally used and frequently abused UV-Visible technique.

Today, derivative spectroscopy allows a fresh look to be taken at previously unresolved or partially resolved UV-Visible problems.

Obviously, the field of application for derivative techniques is extremely wide, thus only a few selected examples are presented here.

The study of steroids, using 1st derivative spectroscopy, has been reported by Olson and Alway [9-Part I], who were, for example, able to identify 6 peaks in the spectrum of testosterone using the zero point crossings. Using the 2nd derivative mode (scan speed of 100 nm/minute and 1 nm S.B.W), the broad, rather featureless zero order spectrum of testosterone (Fluka, purum grade) dissolved in dioxane (Fluka, spectroscopic grade) shows 6 quite distinctive negative peaks in the 280-380 nm region (Figure 32).

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	Trace A		UPO	erator	Sample	()	0
	Mode Ab	5	SBI	A COMP 1.0	700 0 200		C
	Ord Max/Mi	n 1.000	0.000 WL	Max/Min Com/	300.0 200		
	Speed (nm/	min) 100	B*.	line Corr			
	24 Pea	ks, threshol	6 0.010				
	Min 379.2	nm; 0.011	Min 373.4	nm, 0.027			
	Min 368.6	nm, 0.066	Min 350.6	nm, 0.268			
	Min 343.9	nm, 0,365	Min 340.6	NB; 0.371			
	Min 336.8	nm, 0.437	Min 332.3	NM. 0.487			
	Max 329.7	nm, 0.501	Max 317.6	nm, 0.480			
	Max 315.0	nm, 9.464	Max 313.8	nm, 0.433			
	Min 310.7	nm, 0.418	Max 310.5	NM, 0.430			
	Nax 309.3	9 nm - 0.417	Max 308.4	NH . 402			
	Max 304.5	nm, 0.368	Max 302.9	nm, 0.347			
	Max 296.	' nm, 0,297	Max 295.2	nm, 0.288			
	Max 293.6	na, 0.274	Max 291.2	nm, 0.263			
	Max 288.	nm, 0.249	Min 283.5	nm, 0.237			
	Trace B		0P	erator	Sample		
	Mode 2	ND Der	SB	W (nm) 1.0	Smooth	ing (sec)	1
0.005	Ord Max/M	in 0.005	40.005 WL	, Max/Min (nm)	380.0 28	0.0	
	Speed (nm	(min) 100	8'	line Corr			
	3 Pe	aks, thresho	ld 0.005				
2 N.D	Max 353.	5 nm, 0.003	Max 337.3	na, 0.002			
Der	Min 330.	2 nm, -0.004					
1							
0 003					2		
0.000				/	1		
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-6.005	700 0	328 P	34	9.8	360.0	nm. 3	80.0
280.0	200.0	320.0					



The DMS 200 zoom facility was used to locate the wavelengths of the 5 less intense peaks. The agreement between the two sets of peak wavelengths is acceptable, as the solvents used may have been different:

Negative Peak	Zero Order Crossing
(2nd Derivative)	(1st Derivative — Ref. 9)
361.8 nm	361 nm
344.7 nm	344 nm
330.2 nm	332 nm
317.2 nm	324 nm
305.6 nm	310 nm
ca. 293.0 nm	287 nm

and indicates the possibilities of derivative spectroscopic characterization of steroids with modern UV-Visible instruments.

An interesting problem in the field of inorganic chemistry is the determination of the exact number and location of peaks in the UV-Visible spectrum of the uranyl ion (UO_2^{++}) in the 330 to 500 nm region. Using derivative techniques, the 19 bands can be very easily resolved and their positions established with some accuracy (Figure 33). Again the zoom facility can be used for the exact location of weaker intensity peaks.



Figure 33



Figure 34

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The long wavelength peaks are at 327.5 nm, 321.3 nm and 315.5 nm. However, the 2nd derivative spectrum, at a scanning speed of 20 nm/min, indicates the presence of additional peaks (Figure 35).



Figure 35

Similarly, the so-called 258 nm peak can be resolved by 2nd derivative spectroscopy into at least 6 quite distinct individual peaks (Figure 36), located at 267.3 nm, 260.3 nm, 256.2 nm, 253.8 nm, 250.2 nm and 247.5 nm.



Figure 36

In the spectrum of the isomeric diazine, pyrimidine, the two peak groups are shifted towards shorter wavelengths and show less resolved fine structure in the zero order UV spectrum (Figure 37).

Trac	:e	A		,			0₽	erat	or		Sa	mple		
Mode	1	Abs	5				SBI	4 Cn	a)	0.2	Sm	oothing	(sec)	6
Ord	Max	/Mi)	h	3.000	0.000		HL	Max	/Min	(nm)	400.0	210.0		
Spee	e C	næ/a	in)	50			81	line	Cor	r				
	32	Peal	ks,	threshol	d 0.2	88								
Min	35	8.7	08.	-0.004	Min	316	5.4	0.002	0.	322				
Min	30	B.4	080	0.683	Max	298	3.1	0.00 -	8.	988				
Max	28	8.0	082	0.767	Min	262	2.9		0.	487				
Min	25	1.1	DB	2.453	Min	256	3.1	0.0	2.	536				
Min	24	3.6		2.588	Max	245	3.1		2.	838				
Min	24	7.3	Dilla	2.603	Max	245	5.8	0.0.	2.	855				
Min	24	5.4	0.0	2.618	Max	240	1.8	0.	2.5	850				
Min	24	3.9	0.8.	2.641	May	242	2 6		2	979				
Min	24	2.3		2.638	May	241	2	0.0.	2	919				
Min	24	1.2		2.666	May	241		DBA	2	970				
Max	239	2.7	0.00	2.874	Min	275	t 0	0.00	2	667				
Hav	23	5.4		2 897	May	233	1 0	1100 2	2.1	200				
Hav	27	4		2 513	Misio	200	1 7	0.00.2		77				
Nav	220	5 T		2.313	Mag	227		1108 2	4.	340				
Mag	22	. 0	1100 2	1 641	Hit.	221	- 4	1948 2	1.1	23				
FT4LX	223	1.0	F1 00 2	1.041	rit n	221	17	8-00 ×	1.1	¥∠b				
U I U	- 41		日間・	1.602	ITAX .	212	. 4	BB +	1.5	126				





Again, however, it is possible to resolve these peaks into their component peaks with the derivative technique. The effect of scanning speed on the resolution is quite dramatically illustrated in Figure 38 and Figure 39, the latter, at 20 nm/min scanning speed, showing resolved fine structure.







Figure 39

The corresponding 4th derivative spectrum (Figure 40), although enhancing peak resolution even more, shows more noise and requires greater care in its interpretation.

Trac	e A				0p	erat	or		Sa	mele		
Mode	- 4TI	H De	r		SBI	A Ch	n) (3.2	Sa	oothing	(sec)	1
Ord	Max/Mi	n	0.006	-0.006	WL	Max	/Min ((nm)	340.0	260.0		
Sree	d (nmi/i	nin)	20		B' .	line	Corr					
	26 Peal	ks,	thresho	ld 0.0	85							
Min	325.7	Ω.82. P	-0.002	Max	323.7	0.812	0.00	94				
Min	322.5	D#2>	-0.002	Max	317.0	DM+	0.00	34				
Min	315.4		-0.005	Max	313.6	0.81	0.00	34				
Min	312.0	0.00 -	-0.003	Max	310.6	0.00 +	0.00	34				
Min	309.1	082	-0.005	Min	307.9	0.00 -	0.00	90				
Max	307.1	0.00 2	0.087	Min	305.5	0.80 +	-0.00	95				
Max	384.7	0.002	0.004	Min	304.0	0.00	-0.00	31				
Max	303.3	nm,	0.004	Min	302.7	0.00	-0.00	95				
Max	301.0	nm,	0.004	Min	300.1	D M /	-0.00	96				
Max	298.2	0.81 -	0.004	Min	296.2	0.0.	-8.96	34				
Max	295.0	087	0.002	Min -	293.6	0.8.1	-0.00	4				
Max	292.8	0.00 -	0.003	Min	291.0	0.00	-0.00	14				
Max	289.9		0.003	Min	272.1	0.00	-0.00	33				



Figure 40

Similarly, the low UV peak envelope is readily resolved into the major peaks in the 2nd derivative spectrum (Figure 41).

Trad Mode Ord	e A Abs Max/Min d (om/min)	1.500	0.000	Operato SBW (na WL Max/	ir 5 0.2 Min (nm)	Sample Smoothing (sec) 0 260.0 210.0
OF 44	32 Peaks	threshol	d 0.0	50		
Min	259.7 nm	0.076	Min	256.5 nm.	0.126	
Min	255.0 nm.	8.192	Min	254.2 nm.	0.244	Trace B Operator Sample
Min	252.5 nm.	0.419	Min	251.1 nm.	0.567	Mode 2ND Der SBN (nm) 0.2 Smoothing (sec)
Min	250.5 nm,	0.620	Min	249.8 nm-	0.668	Ord Max/Min 0.040 -0.040 HL Max/Min (nm) 260.0 210.0
Min	249.8 nm,	0.715	Min	248.0 nm,	0.764	Speed (nm/min) 20
Min	247.1 nm-	0.841	Min	246.3 nm.	0.932	23 Peaks, threshold 0.005
Max	244.2 nm.	1.074	Min	241.9 nm,	0.998	Min 258,4 nm, 0.002 Max 255.0 nm, 0.012
Max	239.9 nm,	1.054	Max	238.5 nm.	1.004	Max 253.6 nm, 0.010 Min 249.6 nm, -0.012
Max	237.9 nm,	0.953	Max	237.8 nm.	0.864	Max 247.3 nm, 0.015 Min 244.6 nm, -0.031
Max	236.2 nm,	0.814	Max	234.8 nm+	0.771	Max 242.0 nm, 0.014 Min 238.4 nm, -0.024
Max	233.5 nm,	0.717	Max	232.4 nm;	0.639	Max 236.1 nm, 0.019 Min 233.4 nm, -0.015
Max	231.5 nm,	0.552	Nax	230.5 nm,	0.503	Max 231.4 nm, 0.011 Min 228.1 nm, -0.008
Max	228.9 nm,	0.452	Max	227.7 nm.	0.399	Max 226.3 nm, 0.008 Min 223.1 nm, -0.001
flax	226.5 nm.	0.344	Min	222.0 nm>	0.282	Max 221.8 nm, 0.005 Min 218.6 nm, -8.001
Min	218.3 nm,	0.330	Min	215.8 nm,	0.385	Max 215.8 nm, 0.004 Min 214.8 nm, -0.007
Min	213.7 nm	0.432	Min	211.8 nm.	0.479	Max 213.8 nm, 0.005 Min 213.2 nm, -0.001
						Max 212.5 nm, 0.004 Min 211.8 nm, -0.003
						Max 211.1 nm, 0.003





In the third diazine isomer, pyridazine, the less intense peak envelope is quite significantly shifted to longer wavelengths (Figure 42).



Figure 42

The resolution obtained in the 2nd order derivative spectrum (Figure 43) hints at a very complex peak structure, which is further resolved in the 4th order derivative spectrum (Figure 44), where the less intense peak positions can be obtained by the 'zoom' facility.









The short wavelength peak envelope can be likewise characterized by going to higher derivatives (Figure 45 and Figure 46).





Trac	e A				0Pe	erati	or		San	arle	(
uode	2N	n ne	r		281	4 <u>(</u> NI	R 2	0.2	380	0010109	(sec)	1
Ord	Max/Mi	n	0.100	-0.100	WL	Max	∕Min	(nm)	270.0	210.0		
Spee	d (nm/	min>	50		81	line	Corr					
	16 Pea	ks,	threshol	d 0.00	35							
Max	259.5	DM+	0.096	Min	256.1	082	-0.1	03				
Max	253.5	nm.	0.010	Min	251.1	DB2	-0.0	48				
Max	246.6	082	0.042	Min	244.2	087	-0.0	83				
Max	241.7	nm -	0.039	Min	239.2	002	-0.0	78				
Max	236.3	085	0.065	Min	233.8	08.2	-0.0	48				
Max	231.3	N #12	0.051	Min	228.7	₽₩ ≥	-0.0	11				
Max	226.3	n⊪,	0.022	Min	223.9	0.00 2	0.0	02				
Min	219.4	D∰⇒	0.008	Max	212.0	ាតា »	0.0	24				

Trace B	Operator	Sample
Mode 4TH Der	SBW (nm) 0.2	Smoothins (sec) 1
Ord Max/Min 0.040 -0.0	40 – WL Max/Min (nm)	270.0 210.0
Speed (nm/min) 50	B'line Corr	
22 Peaks, threshold 0	.005	
Max 262.0 nm, 0.003 Mi	n 259.0 nm, -0.016	
Max 256.5 nm, 0.024 Ma	x 255.7 nm, 8.019	
Min 253.9 nm, -0.016 Mi	n 252.3 nm, -0.001	
Max 251.3 nm, 0.013 Mi	n 250.2 nm, -0.004	
Min 249.1 nm, -0.000 Ma	x 248.2 nm, 0.012	
Min 246.6 nm, -0.022 Ma	x 244.2 nm, 0.030	
Min 242.1 nm, -0.023 Ma	x 239.1 nm, 0.019	
Min 236.4 nm, -0.023 Ma	x 233.8 nm, 0.020	
Min 231.4 nm, -0.014 Ma	x 223.9 nm, 0.008	
Min 226.4 nm, -0.005 Ma	x 224.0 nm, 0.003	
Max 214.1 nm, 0.002 Mi	n 211.4 nm, -0.008	



Figure 46

These three examples clearly illustrate the potential usefulness of derivative techniques for the re-investigation of the UV-Visible spectra of many compounds, and not only members of homologous or isomeric series.

Quantitative Determination of Trace Compounds

The ability to locate hidden peaks in a spectrum of overlapping peaks makes the derivative technique of particular interest for the quantitation of trace components in complex matrices.

For example, the spectrum of caffeine (1,3,7-trimethylxanthine) in water shows two fairly broad peaks at about 273 nm and 204 nm, together with a prominent shoulder between these two peaks (Figure 47).





The 2nd derivative shows this shoulder to be at about 231 nm.

Caffeine occurs widely in natural products as well as in some commercial products such as, for example, COCA-COLA and PEPSI-COLA.

The UV zero order spectra and the derivative spectra of these COLA's (degassed and diluted 50 times) bear a close resemblance to the pure caffeine spectra. Thus, in the COCA-COLA spectra (Figure 48) and in the PEPSI-COLA spectra (Figure 49) the three peaks occur at about 276-278 nm, 230 nm and 205 nm.







Figure 49

Using the method of standard additions and measuring the 2nd derivative peak amplitudes, D_L, of the 273 nm peak (Figure 50) linear calibrations were obtained (Figure 51) which gave calculated values for caffeine of about 100 mg/l in PEPSI-COLA and 300 mg/l in COCA-COLA.









Of course, the other two wavelength peak amplitudes, as well as the zero-line, D_z , and short wavelength side, D_s , amplitudes may also be used for calibrations and concentration calculations, if they are appropriately linear over the concentration range of interest.

This, by no means exhaustive investigation, shows the potential usefulness of derivative peak amplitudes for the quantitation of both major and minor constituents in natural and commercial materials.

As discussed in Part I, higher derivative techniques can be used to advantage to eliminate completely or at least to minimize undesirable, interfering background which may be present due to matrix absorption or scattering.

For example, the spectrum of a dilute acid solution of the lanthanide, holmium, shows no significant background in the region 235-260 nm, so that the holmium absorption peak at about 241 nm can be readily quantified, with or without the aid of derivative spectroscopy (Figure 52).





However, the spectrum of a holmium glass filter in this region shows a severely increasing background towards shorter wavelengths. (Figure 53)





This background is completely eliminated already in the 2nd order derivative spectrum, so that in this case there is no particular need to go to the 4th or higher order derivatives in order to quantify the peak at 241 nm.

In the author's opinion, in background absorption or scattering situations the use of derivative spectroscopy is very often much simpler and more safe to use than the more commonly used 3- or 2-point correction techniques, which are based on the assumption that over a short wavelength range the change in background absorption is linear.

Characterization of Commercial Materials and Natural Substances

The unambiguous characterization of most natural products and commercial materials by the UV-Visible spectrophotometric technique alone is seldom completely successful, because most substances exhibit rather broad, featureless, non-specific absorption bands, particularly in the UV region. However, higher derivative spectroscopy does offer greater possibilities to obtain more characteristic, archivable 'finger-print' spectra of many substances.

In order to highlight the potential of higher order derivative spectroscopic techniques for finger-printing, several readily available materials were measured. It should be noted, however, that in this preliminary illustrative survey no exhaustive optimization of the instrument operating parameters was attempted.

A sample of a commercially available olive oil, dissolved in chloroform, shows in the zero order spectrum (Figure 54) a single peak at 243 nm and several broad shoulders towards longer wavelengths.



Figure 54

These shoulders are easily resolved in the 2nd order derivative spectrum into four quite distinctive peaks (Figure 55), which could be useful for either characterization or comparison purposes.





A commercial material such as a perfume or aftershave lotion is, of course, a very complex mixture of substances with extensive overlapping of absorption bands, which only the manufacturer is able to characterize easily. Nevertheless, it is possible to resolve some of the overlapping peaks with derivative techniques, as shown in the following spectrum of an aftershave lotion (Figure 56).





The study of tea infusions with derivative spectroscopy gives certainly more information than the zero order spectra. A 'normal' tea spectrum (Figure 56) shows essentially two peaks only. However, these peaks are shown to have a much more complex structure in higher order derivative spectra (Figure 57).





A camomile tea infusion shows very broad peaks which can be resolved into a number of sharp peaks (Figure 58).





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Transparent, solid materials, such as plastic films, also often exhibit rather broad absorption peaks in their zero order spectra, but these can be readily resolved with higher order derivative spectroscopy (Figure 60).





Alcoholic beverages often exhibit rather featureless spectra which can however be considerably enhanced by the use of derivative techniques.

An intensity colorful liqueur, such as BLUE CURACAO, exhibits three peaks in the UV-Visible region (Figure 61) each of which can be readily resolved into several peaks in the derivative spectrum.



Figure 61

Similarly, the digestive liquers FERNET BRANCA and MENTA show virtually identical, poorly resolved zero order spectra which are resolved into several quite distinctive peaks with derivative spectroscopy (Figure 62 and Figure 63).





Trace A Mode Abs Ord Max/Min 3.000 Speed (nm/min) 100 40 Peaks, threshold Min 308.4 nm, 0.323 Min 310.4 nm, 1.233 Max 299.0 nm, 1.392 Max 285.8 nm, 1.261 Min 250.0 nm, 1.506 Min 237.3 nm, 1.752 Min 220.8 nm, 2.347 Min 217.8 nm, 2.443 Max 213.7 nm, 2.573 Max 210.9 nm, 2.573 Max 210.9 nm, 2.573 Min 199.3 nm, 2.675 Min 199.4 nm, 2.675 Min 193.4 nm, 2.954 Min 192.5 nm, 2.954	Operator SBW (mm) 1.0 0.000 WL Max/Min (mm) 0.000 WL Max/Min (nm) 0.000 WL Max/Min (nm) 0.000 WL Max/Min (nm) 0.000 ML Max/Min (nm)	Sample Smoothing (sec) 0 398.0 190.0 Trace B Mode 2ND Der Ord Max/Min 0.030 Speed (nm/min) 100 13 Peaks, thresho Max 348.5 nm, 0.002 Min 279.2 nm, -0.001 Min 252.3 nm, -0.029 Min 243.4 nm, -0.000 Min 225.4 nm, -0.000 Min 209.7 nm, -0.066	Dperator SBW (nm) 1.0 -0.030 WL Max/Min (nm) 1d 0.005 Min 304.6 nm, -0.006 Max 258.4 nm, 0.623 Max 246.5 nm, 0.005 Max 238.4 nm, 0.008 Max 220.4 nm, 0.002 Max 199.8 nm, 0.618	Sample Smoothing (sec) 390.0 190.0
0.030 M				
2ND Der	www	٨		
0.013 M	~ ~			
0.006	A			В
-0.006			X	
-0.018	-			244 Jul
				A
-0.030				700 0
190.0	230.0	270.0 3	10.0 330.0	nm. 370.0

Figure 63

These examples illustrate the huge potential which exists for the application of the derivative technique to the characterization of both natural substances as well as commercial substances.

Conclusion

It is hoped that in this discussion and from the number of illustrative examples presented the usefulness of derivative UV-Visible spectrophotometric techniques for a wide variety of areas of application has been highlighted.

A final word of caution must be introduced, namely that derivative spectra cannot provide any extra information that is not already present in the original zero order absorbance spectra. Derivative spectroscopy simply presents the information in other, visually more easily interpretable format. Thus, it cannot be stressed too highly that the quality and usefulness of derivative spectra will be completely dependent on the intrinsic performance of the UV-Visible spectrophotometer used, even when operated under the optimum measurement conditions.

References

- [122] A.F. Fell and J.G. Allan; Anal. Proc. (London) 18,291 (1981)
- [123] A. Lopez, P. Mazzeo, M.G. Quaglia and F. Segnalini; Il Farmaco-Ed. Pr. 37 (11) 371 (1982)
- [124] A.F. Fell, H.P. Scott, R. Gill and A.C. Moffat; Anal. Proc. (London) 20, 173 (1983)
- [125] R.F. Haines-Nutt and P. Adams; Anal. Proc. (London) 21, 241 (1984)
- [126] M.E. Abdel-Hamid, M.M. Abdel-Khalek and M.S. Mahrous; Anal. Lett. 17 (B 12) 1353 (1984)
- [127] M.A. Korany, A.M. Wahbi, M.A. Elsayed and S. Mandour; Anal. Lett. 17 (B 12) 1373 (1984)
- [128] S. Honda, T. Konishi and H. Chiba; Anal. Chem. 56, 2352 (1984)
- [129] M.A. Korany, A.M. Wahbi, M.A. Elsayed and S. Mandour; Il Farmaco, Ed.Prat., 39 (7) 243 (1984)
- [130] K. Kitamura, M. Takagi and K. Hozumi; Chem. Pharm. Bull. 32, (4) 1484 (1984)
- [131] Y. Pang and Y. Cui; Yaowu Fenxi Zazhi, 4 (1) 25 (1984)
- [132] J. Xu and R. Gan; Yaowu Fenxi Zazhi, 4 (2) 124 (1984)
- [133] Q. Yang, Y. Meng and G. Zhang; Yaowu Fenxi Zazhi, 4 (3) 148 (1984)
- [134] J. Zhang, Z. Deng and H. Zeng; Yaowu Fenxi Zazhi, 4 (3) 157 (1984)
- [135] Y. Fuki, M. Matsubara, S. Takahashi and K. Matsubara; J. Anal. Toxicol. 8 (6) 277 (1984)
- [136] T.J. Siek and F. Rieders; J. Forensic Sci. 29 (1) 39 (1984)
- [137] M.A. Korany, A.M. Wahbi, S. Mandour and M.A. Elsayed; Anal. Lett. 18 (B 1) 21 (1985)
- [138] L. Sun, Q. Yang and R. Yue; Yaowu Fenxi Zazhi, 5 (1) 23 (1985)
- [139] A. Bettero and P. Bollettin; Anal. Chim. (Roma) 75 (7-8) 351 (1985)
- [140] F.A. El-Yazbi and M.H. Barary; Anal. Lett. 18 (B 5) 629 (1985)
- [141] A. Lezerovich; J. Am. Oil chem. Soc. 62 (5) 883 (1985)
- [142] M.E. Abdel-Hamid, M.H. Barary, M.A. Korany and E.M. Hassan; Sci. Pharm. 53 (2) 105 (1985)
- [143] X. Yuan; Yaowu Fenxi Zazhi, 5 (2) 120 (1985)
- [144] M.A. Korany, F.A. El-Yazbi, O. Abdel-Razak, and M.A. Elsayed; Pharm. Weekbl. Sci. Ed. 7 (4) 163 (1985)
- [145] Tsung-Li Kuo; Clin. Chem. 32 (2) 337 (1986)
- [146] G.E. James; Hewlett-Packard Journal, Feb. 1980, 5.
- [147] A.A. Kucher, N.S. Poluektov, V.T. Mishchenko, N. Aleksandrova; Zavod. Lab. 49 (10) 11 (1983)
- [148] R. Qu and Z. Xue; Fenxi Huaxue, 12 (6) 516 (1984)
- [149] P. Pokrowsky and W. Herrmann; Opt. Eng. 23 (1) 88 (1984)
- [150] P.K. Spitsin; Zavod. Lab. 51 (3) 16 (1985)
- [151] L. Lepine, R. Gilbert and G. Belanger; Anal. Chem. 58 (6) 1152 (1986)
- [152] R.L. Sharma, H.B. Singh, M. Satake; Analyst, 111, 551 (1986)
- [153] K. Nagashima, Xue-Xin Qian, and S. Suzuki; Analyst, 111, 771 (1986)
- [154] P. Gans; Anal. Proc. (London) 17, 133 (1980)
- [155] A.F. Fell; Anal. Proc. (London) 17, 266 (1980)
- [156] T.C. O'Haver; Anal. Proc. (London) 19, 22 (1982)
- [157] K. Yan; Huaxue Tongbao, 8, 16 (1984)
- [158] M.A. Korany, A.A. Seif El-Din and N.A. Abdel-Salam; Anal. Lett. 17, (A 6) 483 (1984)
- [159] H. Kadin; Anal. Lett. 17, (A 11) 1245 (1984)
- [160] J. Gartzke, K-D. Nolte and K. Berka; Jena Rev. 4, 170 (1984)
- [161] A.R. Hawthorne, S.A. Morris, R.L. Moody and R.B. Gammage; J. Environ. Sci. Health, Part A, A 19 (3) 253 (1984)

- [162] A. Etournaud and J-D. Aubort; Mitt. Geb. Lebensmittelunters. Hyg. 75 (2) 221 (1984)
- [163] R. Riedl, W. Luf and E. Brandl; Z. Lebensm.-Unters.-Forsch. 179 (5) 394 (1984)
- [164] G. Gauglitz, T. Klink and A. Lorch; Fresenius Z. Anal. Chem. 319 (4) 364 (1984)
- [165] A. Werle-Wilczynska, D. Ciecierska-Stoklosa, K. Gorczynska and M. Gluzinska; J. Mol. Struct. 115, 185 (1984)
- [166] P. Gans and J.B. Gill; Appl. Spectrosc. 38 (3) 370 (1984)
- [167] M.J. Scott; Beckman Technical Information, T-1573-UV-84-11
- [168] Y.R. Tahboub and H.L. Pardue; Anal. Chem. 57, 38 (1985)
- [169] P. Chen, Q. Luo, Y. Zeng; Guangpuxue Yu Guangpu Fenxi, 5 (2) 5 (1985)
- [170] K. Kitamura and K. Hozumi; Anal. Chim. Acta, 172, 111 (1985)
- [171] L. Dixit, S. Ram; Appl. Spectrosc. Rev. 21 (4) 311 (1985)
- [172] V.A. Peril'ev, V.T. Mishchenko, N.S. Poluektov; Zh. Anal. Khim., 40 (8) 1349 (1985)
- [173] L. Dixit, S. Ram, R.B. Gupta, H.C. Chandola and P. Kumar; Analyst, 111, 101 (1986)
- [174] L. Meal; Anal. Chem. 58, 834 (1986)
- [175] J.A. Howell and L.G. Hargis; Anal. Chem. 58 (5) 113 R (1986)
- [176] A-M.M.Wahbi, M.A. Abounassif and H.M.G. Al-Kahtani; Analyst, 111, 777 (1986)
- [177] Varian Brochure, 'Varian's DMS 200 an Advanced UV-VIS Spectrophotometer.'
- [178] C. Burgess and A. Knowles; 'Techniques in Visible and Ultraviolet Spectrometry Volume 1 Standards in Absorption Spectrometry — Ultraviolet Spectrometry Group' 7, 111. Publ. Chapman and Hall Ltd. 1981
- [179] Varian 'Optimum Parameters for Spectrophotometry' OPT-720 (1973)
- [180] R.A. Morton; 'Biochemical Spectroscopy', Vol. 1 p. 260, 261, Adam Hilger (1975)